FIST-GT: A tool for multidimensional spatial transcriptomics data imputation via graph-regularized tensor completion

Thomas Atkins Tianci Song Rui Kuang Department of Computer Science and Engineering, University of Minnesota January 13, 2022

Background

Spatial transcriptomics (ST) is a highly promising new technology for measuring gene expression across a tissue section that captures spatial heterogeneity of the whole transcriptome. The technology captures two dimensional gene expression profile from tissue sections. Furthermore, parallel tissue slices can be combined to create a three dimensional gene expression map. However, data from current experimental technologies is significantly zero-inflated due to low capture efficiency,

Pipeline

Our method takes 3D data as a table of genes by spots (continuous in 3D space), converts it to a discrete tensor representation, imputes this tensor, and then interpolates it back into the continuous spot data.



Results (cont.)

Additionally, we measure performance using the following three metrics:

Mean absolute error (MAE) = ¹/_n ∑_i |T_i - Î_i.
Symmetric mean absolute percentage error (SMAPE) = ¹/_n ∑_i |T_i - Î_i.
Coefficient of determination (R²): 1 - (∑_i(T_i - Î_i)²) (∑_i(T_i - T̄)²)⁻¹

necessitating a means of data reconstruction.

Methods

Data

Let $\mathcal{T} \in \mathbb{R}^{n_p \times n_1 \times \ldots \times n_N}_+$ be the observed gene expression tensor, where n_p denotes the number of genes, and n_1, \ldots, n_N represent the size of the spatial dimensions. We approximate \mathcal{T} with $\hat{\mathcal{T}}$ where $\hat{\mathcal{T}}$ is represented in the Canonical Polyadic Decomposition (CPD) form, so $\hat{\mathcal{T}} = \sum_{i=1}^r \bigotimes_{i=1}^N [\hat{A}_p]_{:,i}$.

Now, we let the following be our objective function, similar to [1]:

 $\min_{\hat{A}_{i},i\in\{x_{1},...,x_{n},p\}}\frac{1}{2}||\mathcal{M}\circledast(\mathcal{T}-\hat{\mathcal{T}})||_{\mathcal{F}}^{2}+\frac{\lambda}{2}\mathbf{vec}(\hat{\mathcal{T}})^{T}\mathfrak{L}(x_{1},...,x_{n},p)\mathbf{vec}(\hat{\mathcal{T}})$

where \mathcal{M} is a binary mask tensor of the observed values and $\mathfrak{L}(x_1, ..., x_n, p)$ is the graph Laplacian of the Cartesian product of G_{x_1} , ..., G_{x_N} , and G_p , where G_{x_i} is a spatial chain graph of x_i , and G_p is a protein-protein interaction network. To minimize our objective function we utilize a multiplicative update rule (omitted here for brevity).





We test the method on three 3D gene expression datasets. The first measures gene expression in the developing human heart at 6.5 PCW (DHH). [2] The dataset was created by mapping 9 tissue slices sequenced using ST into one tissue atlas. The second dataset is an expression atlas of the adult mouse brain (AMB). [3] This dataset was similarly prepared through ST sequencing of parallel 2-dimensional sequences, assembled to form a three-dimensional dataset. The third is an atlas of developmental genes in a stage 5/6 *Drosophila* embryo (DME). [4] Unlike this other two datasets, this dataset was obtained through fluorescent antisense RNA imaging. The three datasets are visualized below, colored based on expression of a sample spatially variable genes.



Results

We compare our model (FIST-GT) to a spatial nearest-neighbor model (SNN) using 5-fold cross validation. Here, we plot the cumulative distribution function (CDF) of absolute errors for the two methods, and see that the error CDF of FIST-GT is generally less than the error CDF of SNN.



Figure: Comparison of MAE, SMAPE, and R^2 for FIST-GT and SpatialNN on 5-fold cross validation of three datasets.

From the chart above, we see that FIST-GT clearly outperforms SNN for almost every metric on nearly every dataset (the exception being SMAPE on the DME dataset, which is approximately equal for both methods).

Conclusions and Future Directions

Here we have shown that FIST imputes three-dimensional spatial expression data from a variety of datasets more accurately than a spatial nearest-neighbor model. The heterogeneity of the datasets tested demonstrates that the method is widely applicable. Future work will measure errors between the original unbinned data and output interpolated data to ensure the method's real-world utility.

References



Additionally, the number of genes obtained in every dataset can be found in the table below:

DatasetMethod n_p DHHStacked ST13850AMBStacked ST14035DMEmRNA Imaging84

Figure: Cumulative absolute error distributions for both models on all datasets.A one-sided paired Wilcoxon signed-rank test on the error distributionsof the two methods produced the following:Dataset StatisticpDHH $6.4 \cdot 10^{11} < 0.001$ AMB $4.8 \cdot 10^{13}$ OME $1.6 \cdot 10^9$ 0.79

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